This article was downloaded by: [University of California, San Diego]

On: 15 August 2012, At: 23:21 Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH,

UK



Molecular Crystals and Liquid Crystals Science and Technology. Section A. Molecular Crystals and Liquid Crystals

Publication details, including instructions for authors and subscription information: http://www.tandfonline.com/loi/gmcl19

Nucleobase Mapping of Self-Assembled Monolayers by Chemical Force Microscopy

Hiroshi Sunami ^a , Kuniharu Ijiro ^a , Olaf Karthaus ^a , Stephan Kraemer ^b , Silvia Mittler ^b , Wolfgang Knoll ^b & Masatsugu Simomura ^a

Version of record first published: 24 Sep 2006

To cite this article: Hiroshi Sunami, Kuniharu Ijiro, Olaf Karthaus, Stephan Kraemer, Silvia Mittler, Wolfgang Knoll & Masatsugu Simomura (2001): Nucleobase Mapping of Self-Assembled Monolayers by Chemical Force Microscopy, Molecular Crystals and Liquid Crystals Science and Technology. Section A. Molecular Crystals and Liquid Crystals, 371:1, 151-154

To link to this article: http://dx.doi.org/10.1080/10587250108024710

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.tandfonline.com/page/terms-and-conditions

^a Research Institute for Electronic Science, Hokkaido University, Sapporo, 060-0812, Japan

^b Max-Planck-Institute for Polymer Research, Mainz, Germany

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae, and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand, or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

Nucleobase Mapping of Self-Assembled Monolayers by Chemical Force Microscopy

HIROSHI SUNAMI¹, KUNIHARU IJIRO¹, OLAF KARTHAUS¹, STEPHAN KRAEMER², SILVIA MITTLER², WOLFGANG KNOLL² and MASATSUGU SIMOMURA¹*

¹Research Institute for Electronic Science, Hokkaido University, Sapporo 060-0812, Japan and ²Max-Planck-Institute for Polymer Research, Mainz, Germany

In order to measure complementary hydrogen bonding between nucleobases, disulfide derivatives of thymine and adenine were newly synthesized. Surface Plasmon Resonance; SPR confirmed the formation of SAMs on Au. SPR shows that SAMs of nucleobase derivatives were completely formed within 60 minutes. Thickness of the adenine SAM and the thymine SAM is nearly the same. AFM tip was modified with the thymine SAM. The adenine SAM-modified Au electrode was clearly observed in an adhesion force map. The adhesion force between complementary nucleobases is larger than that of the non-complementary combination.

<u>Keywords</u>: chemical force microscopy; molecular recognition; force mapping; self-assembled monolayers; hydrogen bond; DNA

INTRODUCTION

One of the targets of DNA analysis is sequencing nucleobases

of single DNA molecule. Chemical force mapping by a prove-tip modified with nucleobases is a strong analytical method for sequencing of the single DNA molecule. It is known that the hydrogen bonding of a nucleobase to its complement is enhanced in hydrophobic environment at the at an air-water interface¹⁻³. Monolayer of single-alkylated cytosine derivative at the air-water interface shows strong selectivity towards the complementary nucleobases dissolved in the aqueous subphase⁴. Here we propose a method for the binding and detection of nucleobases or monomeric/oligomeric DNA fragments by using a disulfide-modified gold surface as the interface. We herein demonstrate base-pair mapping by using chemical force microscopy.

RESULTS AND DISCUSSION

In order to measure complementary hydrogen bonding between nucleobases, disulfide derivatives of thymine and adenine were synthesized(Figure. 1). Surface Plasmon Resonance; SPR was used to confirm the formation of SAMs on Au. SPR shows that SAMs of nucleobase derivatives were completely formed within 60 minutes when Au substrates were soaked in ethanol solutions of disulfide derivatives (10µM) (Figure. 2). The thickness of the adenine SAM and the thymine SAM are nearly the same.

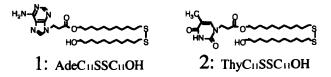
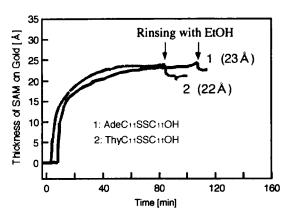


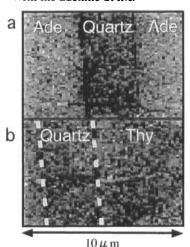
Figure. 1 Chemical formulae of nucleobase disulfide derivatives.



Time courses of SPR due to adsorption of nucleobase disulfide derivatives in EtOH solution on gold at room temperature.

Gold-coated AFM tips and Au electrodes were modified with the adenine or thymine SAM, respectively. Comb-shape patterned Au electrode (width; 10µm, interval; 5µm, length; 2mm, number; 65) deposited on quartz surface was purchased from NTT advanced technology. Tips and electrodes were soaked in 1mM ethanol solution for 24h at room temperature. Adhesion force curves were measured in pure water at 20°C by using OLYMPUS-NV2500 AFM (spring constant; 0.18N/m). The adhesion forces were mapped as 64×64 dots image. The adhesion force maps of the thymine- or adenine-modified patterned Au surfaces by the thymine-modified AFM tip are shown in Figure. 3. Alarge contrast between the quartz surface and the Ademodified gold surface is clearly observed in the force map. While, in the Thy-modified electrode, homogeneous image was observed. Histograms of adhesion forces were obtained from the force-distance curves of 1024 measurement points (Figure. 4). The adhesion force (mean value; 2.01nN) of the complementary adenine-thymine combination is larger than the non-complementary combination (mean value; 1.02nN). Similar results were observed in the adhesion force mapping of patterned Au electrodes when the AFM tip was modified

with the adenine SAM.



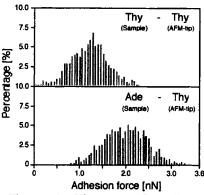


Figure. 4 Histograms for adhesion force obtained from the force-distance curves.

Figure. 3 Mapping of adhesion force between a thymine modified tip and adenine(a) or thymine modified gold micro electrode(b), réspectively. White dots lines indicate the electrode edges observed by the topological imaging of AFM.

Reference

- [1] H. Kitano, H. Ringsdorf, Bull. Chem. Soc. Jpn., 2826, 58 (1985).
- [2] Y. Sasaki, K. Kurihara, T. Kunitake, J. Am. Chem. Soc., 10994, 114 (1992).
- [3] T. Krauch, S. Yu, P. Zubov, Colloids Surf., 383, 57 (1991).
- [4] M. Shimomura et al., J. Am. Chem. Soc., 2341, 119 (1997).